

# The Adrenergic Pathway and Heart Failure

J.R. KEYS\* AND W.J. KOCH†

\*Department of Surgery, Duke University Medical Center, Durham, North Carolina 27710;

†Center for Translational Medicine, Department of Medicine, Thomas Jefferson University, Philadelphia, Pennsylvania 19107

## ABSTRACT

Heart failure represents the endpoint to many triggering cardiovascular pathologies. However, there are molecular and biochemical features that remain common to the failing heart, despite the varying etiologies. Principal among these is heightened activation of the sympathetic nervous system and associated enhancement of adrenergic signaling pathways via the catecholamines, norepinephrine and epinephrine. During heart failure, several hallmark alterations in the adrenergic system contribute to loss of cardiac function. To specifically study these changes in a physiologically relevant setting, we and others have utilized advances in genetically engineered mouse technology. This chapter will discuss the many transgenic and knockout mouse models that have been developed to study the adrenergic system in the normal and failing heart. These models include genetically manipulated alterations of adrenergic receptors, linked heterotrimeric G proteins, and the regulatory G protein-coupled receptor kinases (GRKs). Among the more-interesting information gained from these models is the finding that inhibition of a particular GRK — GRK2 or  $\beta$  adrenergic receptor kinase 1 ( $\beta$ ARK1) — is a potential novel therapeutic strategy to improve function in the setting of heart failure. Furthermore, we will discuss recent transgenic research that proposes an important role for hypertension in the development of heart failure. Overall, genetically engineered mouse models pertaining to this critical myocardial signaling system have provided novel insight into heart function under normal conditions and during states of dysfunction and failure.

## I. Heart Failure and Sympathetic Nervous System Signaling

More than 500,000 new cases of heart failure are reported each year in the United States alone, making it one of the world's most-prolific diseases. The principal function of the heart is to provide enough oxygenated blood to meet the body's metabolic demands through cardiac output. Although the heart is adaptable to many physiological conditions, various etiologies can perturb its function, leading to ventricular dysfunction and ultimately failure. The initial response of the heart to excessive stress is to enlarge morphologically to a state known as cardiac hypertrophy. Classically, cardiac hypertrophy is defined as the physiological response of the heart to an increased workload. This hypertrophy may serve as compensatory and aid in preventing progressive deterioration of cardiac function (Grossman *et al.*, 1975; Chein, 1999). However, often, the stress will

overwhelm the system, sending the hypertrophied heart into failure. As the disease progresses, the heart dilates and thins, becoming too weak to maintain adequate cardiac output. During these hypertrophic and failing processes, there is sustained heightened activation of the renin-angiotensin system and the sympathetic nervous system in an attempt by the body to maintain cardiac output and systemic blood pressure (Esler *et al.*, 1997). Recent evidence suggests that the signaling pathways stimulated during the hypertrophic process, if left unchecked, participate in the pathogenesis of heart failure and may be more important in this disease process than the actual stress placed on the heart (Esposito *et al.*, 2002; Rockman *et al.*, 2002).

The importance of sympathetic activity, via the catecholamines norepinephrine and epinephrine, in heart failure progression and mortality is well established (Cohn *et al.*, 1984). At a cellular level, the catecholamines act upon the heart by binding to the adrenergic receptors (ARs), which are members of the superfamily of proteins known as the G protein-coupled receptors (GPCRs) (Caron and Lefkowitz, 1993). In the heart, norepinephrine principally binds to the  $\alpha_{1B}$ - and  $\beta_1$ AR, while epinephrine is a ligand for both  $\beta_1$ - and  $\beta_2$ AR (Caron and Lefkowitz, 1993). The  $\beta_1$ AR is the most-abundant  $\beta$ AR in the human heart, approaching 75% of the total number of receptors (Brodde, 1993). The  $\beta$ ARs are coupled primarily to the heterotrimeric G protein, Gs, to stimulate adenylyl cyclase activity. This association generates intracellular cyclic adenosine monophosphate (cAMP) and protein kinase A (PKA) activation, which regulate cardiac contractility and heart rate (Bristow *et al.*, 1989).  $\beta_2$ ARs also can couple to the G protein, Gi, which can inhibit adenylyl cyclase activity and stimulate novel mitogen-activated protein kinase (MAPK) pathways in the heart through G $\alpha$  and G $\beta\gamma$  subunits (Rockman *et al.*, 2002). Alternatively, binding of norepinephrine to the  $\alpha_{1B}$ AR elicits phospholipase C (PLC) activity via activation of the G protein, Gq, which is the principal G protein signaling pathway implicated in the hypertrophic response of the heart (Molkentin and Dorn, 2001).

Following agonist occupation of ARs, these GPCRs become substrates for regulation via G protein-coupled receptor kinases (GRKs), which phosphorylate activated receptor (Inglese *et al.*, 1993). This phosphorylation facilitates binding of  $\beta$ -arrestins, which sterically interferes with further coupling to G proteins, thus desensitizing and uncoupling the signal. The principal GRKs involved in intracellular signaling within the heart are GRK2 (or  $\beta$  adrenergic receptor kinase 1,  $\beta$ ARK1), GRK3, and GRK5, all of which have specific GPCR selectivity *in vivo* in the heart (Eckhart *et al.*, 2000). As will be detailed later, GRK activity in the heart appears to play a critical role, especially in heart failure. The generalized signaling pathways, their regulation, and outcomes in the heart are depicted in Figure 1.

Importantly, in human heart failure, chronic activation of the sympathetic nervous system has adverse implications and can accelerate cardiac pathology

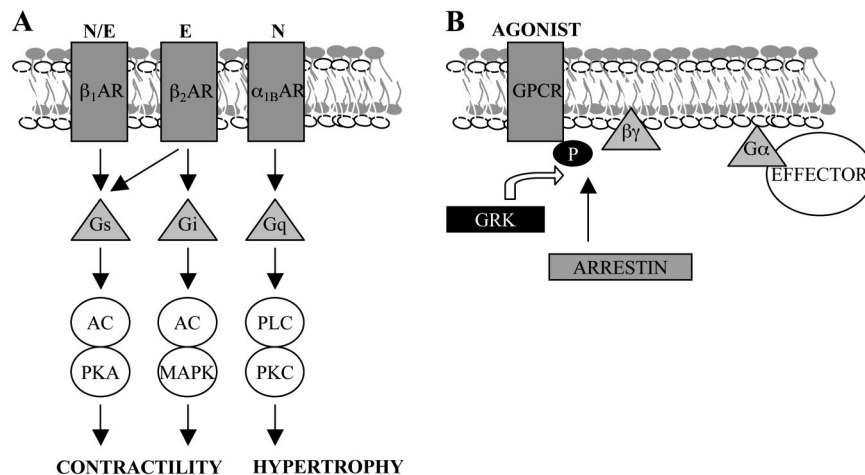


FIG. 1. (A) Schematic representation of the signaling pathways through the various adrenergic receptors (ARs) in the heart. (B) Schematic representation of the process of desensitization of G protein-coupled receptors (GPCRs). Following agonist binding to the GPCR, the associated heterotrimeric G protein splits into  $G\beta\gamma$  and  $G\alpha$ . The  $G\alpha$  elicits a signal through the cell via effector molecules. In addition, the receptor is now a substrate for phosphorylation by the appropriate G protein-coupled receptor kinase (GRK). This phosphorylation facilitates arrestin binding, which sterically interferes with further G protein activation, thus desensitizing and uncoupling the signal. Abbreviations: N, norepinephrine; E, epinephrine; PKA, protein kinase A; MAPK, mitogen-activated protein kinase; PKC, protein kinase C.

(Cohn *et al.*, 1984). Constant stimulation of ARs by catecholamines leads to selective  $\beta_1$ AR downregulation (Bristow *et al.*, 1982,1993; Ungerer *et al.*, 1993). However, both the  $\beta_1$ AR and the  $\beta_2$ AR are markedly uncoupled from their G proteins and effector systems (Bristow *et al.*, 1982,1989). The latter appears to be due to increased levels and GRK activity of BARK1 (Ungerer *et al.*, 1993,1994). In addition,  $G\alpha_i$  is significantly upregulated, to dampen adenylyl cyclase activation (Feldman *et al.*, 1988). Overall, these molecular adrenergic changes in the failing human heart (summarized in Table I) lead to a marked attenuation of cardiac  $\beta$ AR signaling.

Interestingly, genetic polymorphisms in ARs have been identified and may influence individual characteristics of heart failure (Green *et al.*, 1993; Podlowski *et al.*, 2000; Small *et al.*, 2002). For example, individuals with an Ile164 allele for the  $\beta_2$ AR have significantly reduced survival (Green *et al.*, 1993) and lower exercise capacity (Wagoner *et al.*, 2000). In addition, polymorphisms within the  $\beta_1$ AR, combined with a deletion mutant found within the human  $\alpha_{2c}$ AR gene, can act synergistically to increase the risk of heart failure in the black population (Small *et al.*, 2002). These genetic studies, as well as recent heart failure drug

TABLE I  
*Changes in the  $\beta$ AR Signaling Pathway in Human Heart Failure*

| Molecule                 | Change            | References  |
|--------------------------|-------------------|---|
| $\beta_1$ ARs            | ↓, uncoupled      | Bristow <i>et al.</i> , 1982, 1993;<br>Ungerer <i>et al.</i> , 1993, 1994 |
| $\beta_2$ ARs            | NC, uncoupled     | Bristow <i>et al.</i> , 1993; Ungerer <i>et al.</i> , 1993, 1994          |
| $\beta$ ARK1             | ↑ mRNA + activity | Ungerer <i>et al.</i> , 1993  |
| GRK3                     | NC                | Ungerer <i>et al.</i> , 1994  |
| GRK5                     | Not studied       | —   |
| $\beta$ Arrestin 1 and 2 | NC                | Ungerer <i>et al.</i> , 1993  |
| G $\alpha$ i             | ↑                 | Feldman <i>et al.</i> , 1988  |
| G $\alpha$ s             | NC                | Feldman <i>et al.</i> , 1988  |

[Abbreviations: AR, adrenergic receptor; GRK, G protein-coupled receptor kinase; NC, no change.]

trials, have indicated that there is still much to learn about alterations in the  $\beta$ AR system during cardiac failure. This includes recent studies demonstrating significant improvement in survival with  $\beta$ AR antagonist therapy in people with moderate and severe heart failure (Packer *et al.*, 1996,2001). This is contraindicative to the short-term effects of  $\beta$  blockers (Epstein and Braunwald, 1966). The recent use of genetically engineered mice has provided unique experimental models for the study of cardiac adrenergic signaling alterations and the function of the normal and failing heart.

## II. Mouse Models to Study the Cardiac Adrenergic System

### A. LOSS AND GAIN OF AR EXPRESSION

#### 1. The $\beta_1$ AR

Gene-targeted knockout mice with disruption and ablation of the  $\beta_1$ AR gene generally are embryonically lethal. These mice recently have been reviewed in detail (Rohrer, 1998). Although surviving  $\beta_1$ AR knockout mice have normal heart rates, their response to exercise is abrogated (Rohrer *et al.*, 1996). Interestingly, despite the presence of  $\beta_2$ ARs, there was no response to  $\beta$ -agonist stimulation, suggesting that  $\beta_1$ AR is responsible for catecholamine-induced alterations in heart rate in the mouse (Rohrer *et al.*, 1996).

Using the  $\alpha$  myosin heavy chain ( $\alpha$ MHC) promoter to target gene expression to adult ventricular myocardium, mice that overexpress the  $\beta_1$ AR in the heart have been generated (Englhardt *et al.*, 1999). These mice, with 5- to 15-fold overexpression compared to endogenous  $\beta$ AR levels, exhibit a pathology that is consistent with chronic sympathetic stimulation, with a phenotype of dilated cardiomyopathy and heart failure (Englhardt *et al.*, 1999). As will be detailed, this is in striking contrast to phenotypes observed with myocardial  $\beta_2$ AR overexpression.

## 2. The $\beta_2$ AR

In contrast to the  $\beta_1$ AR knockout mice, gene disruption of the  $\beta_2$ AR does not appear to significantly alter cardiac physiology (Rohrer, 1998). This suggests that, under normal conditions, the  $\beta_2$ AR plays no major role in murine cardiac physiology. However, the  $\beta_2$ AR can significantly alter cardiac physiology when overexpressed. Transgenic  $\alpha$ MHC- $\beta_2$ AR mice generated by the Lefkowitz laboratory had greater than 200-fold overexpression of endogenous  $\beta$ ARs. These mice possessed a biochemical and physiological phenotype that mimicked maximal  $\beta$ AR myocardial signaling and function (Milano *et al.*, 1994b). Surprisingly, even though these mice have enhanced heart rates and contractility from a young age, there is minimal pathology present, even in mice greater than 1 year of age (Koch *et al.*, 2000). Furthermore, other transgenic mice with lower levels of overexpression (i.e., 30- to 50-fold) have similar characteristics (Turki *et al.*, 1996). However, Liggett and colleagues (2000) have reported that a “transgene dose-response” for the  $\beta_2$ AR often can have delayed deleterious consequences, similar to what is observed with minimal  $\beta_1$ AR overexpression.

There appears to be significant Gi coupling in  $\alpha$ MHC- $\beta_2$ AR mice, as demonstrated by studies utilizing the Gi inhibitory pertussis toxin (Xiao *et al.*, 1999). These data contribute to recent findings demonstrating that signaling via  $\beta_1$ AR and  $\beta_2$ AR in the heart is fundamentally different (Rockman *et al.*, 2002). The overall positive effects seen with transgenic  $\beta_2$ AR overexpression suggest that the use of genetic engineering to replace lost  $\beta$ ARs with the  $\beta_2$ AR in the failing heart may be therapeutic (Maurice *et al.*, 1999). Consistent with this, we have found that cardiac  $\beta_2$ AR overexpression can “rescue” a mouse model of decompensated hypertrophy and heart failure due to cardiac  $G\alpha_q$  overexpression (Dorn *et al.*, 1999). However,  $\alpha$ MHC- $\beta_2$ AR mice were unable to rescue other mouse models of cardiomyopathy (Rockman *et al.*, 1998b; Freeman *et al.*, 2001). Moreover,  $\beta_2$ AR overexpression leads to functional deterioration of the heart, following induction of pressure overload (Du *et al.*, 2000).

### 3. The $\alpha_{1B}AR$

Two lines of mice have been generated using the  $\alpha$ MHC promoter and the  $\alpha_{1B}AR$ . These mice express either the wild-type receptor or a constitutively active mutant of the  $\alpha_{1B}AR$  (CAM $\alpha_{1B}AR$ ) (Akhter *et al.*, 1997; Milano *et al.*, 1994a). The CAM $\alpha_{1B}AR$  mice exhibit significant myocardial hypertrophy, suggesting that  $\alpha_{1B}AR$  activation can induce cardiac changes independent of hemodynamic influences (Milano *et al.*, 1994a). In contrast, mice overexpressing the wild-type  $\alpha_{1B}AR$  do not develop an increase in heart size, despite displaying some biochemical characteristics of hypertrophy (Akhter *et al.*, 1997). These mice, however, show a reduced tolerance to chronic  $\alpha_{1B}AR$  stimulation, indicating that they are primed for a hypertrophic response (Iaccarino *et al.*, 2001). Interestingly, the  $\alpha$ MHC- $\alpha_{1B}AR$  transgenic mice have a decreased response to  $\beta$ AR stimulation (Akhter *et al.*, 1997), which appears to be mediated via an observed increase in  $\beta$ ARK1 expression and activity (Akhter *et al.*, 1997; Iaccarino *et al.*, 2001) and an activation of the sympathetic nervous system (Iaccarino *et al.*, 2001). Thus, this mouse model has led to an elucidation of molecular cross-talk between the  $\alpha_{1B}AR$  and the  $\beta$ AR systems in the heart. Finally, transgenic mice expressing the  $\alpha_{1B}AR$  under the control of its isogenic promoter exhibit myocardial hypertrophy and have a surprising loss of sympathetic activity (Zusic *et al.*, 2001).

## B. GENETIC ALTERATION OF CARDIAC G PROTEIN EXPRESSION

### 1. $G\alpha_s$

Transgenic mice overexpressing the stimulatory G protein  $G\alpha_s$  in the heart have been generated and characterized (Gaudin *et al.*, 1995). These mice exhibit enhanced responsiveness to catecholamines and develop cardiomyopathy as they age (Geng *et al.*, 1999), in a model reminiscent of human heart failure. Interestingly, the phenotype can be rescued by chronic administration of a  $\beta$ AR blocker (Asai *et al.*, 1999), suggesting that this phenotype, at least partially, mimics chronic sympathetic nervous system activation and enhanced  $\beta$ AR signaling.

### 2. $G\alpha_i$

Targeted disruption of  $G\alpha_i_2$  or  $G\alpha_i_3$  (the major Gi subtypes in myocardium) in mice revealed that there appears to be no significant role for Gi signaling in basal cardiac function or in the response to  $\beta$ AR stimulation in the normal heart (Jain *et al.*, 2001). In contrast to these knockout results, expression of a novel Gi-coupled receptor in the heart resulted in a large decrease in myocardial force, suggesting that defects in the Gi signaling pathway may contribute to the

development of cardiac pathology (Redfern *et al.*, 1999; Baker *et al.*, 2001). These results are consistent with the upregulation of  $G\alpha_i$ , contributing to human heart failure and the uncoupling of the  $\beta$ AR system (Feldman *et al.*, 1988).

### 3. $G\alpha_q$

Dorn and colleagues have described transgenic mice overexpressing  $G\alpha_q$  in the heart (D'Angelo *et al.*, 1997).  $\alpha$ MHC-Gq mice with 4-fold overexpression have cardiac hypertrophy and alterations in all of its molecular markers (D'Angelo *et al.*, 1997). These animals, like the  $\alpha_{1B}$ AR overexpressors, display abrogated  $\alpha$ AR function. Crossbreeding the Gq mice with transgenic mice that had 200-fold overexpression of the  $\beta_2$ AR worsened the Gq phenotype (Dorn *et al.*, 1999). However, a line of  $\beta_2$ AR mice with only 30-fold overexpression of the receptor rescued the cardiac hypertrophy (Dorn *et al.*, 1999), suggesting that selective, controlled  $\beta$ AR enhancement may be beneficial. At higher levels of Gq expression in the transgenic mice, severe heart failure and early death was observed, with a component of increased myocyte apoptosis (Adams *et al.*, 1998).

### 4. $G\alpha_q$ Inhibition

Due to the importance of Gq signaling in the development of cardiac hypertrophy, our laboratory set out to selectively inhibit this pathway in the heart. To achieve this, a specific peptide inhibitor consisting of the last 54 amino acids of the Gq (GqI) was developed and studied (Akhter *et al.*, 1998). This GqI peptide targets the receptor-Gq interface, competitively inhibiting  $G\alpha_q$  activation while not affecting Gs or Gi signaling (Akhter *et al.*, 1998). Transgenic mice expressing the GqI peptide in the heart were shown to have attenuated responses to Gq-coupled receptor stimulation (Akhter *et al.*, 1998). When these animals were subjected to an experimental model of pressure overload cardiac hypertrophy, expression of the GqI peptide in the heart significantly inhibited development of the hypertrophic phenotype (Akhter *et al.*, 1998). Thus, this study identified Gq activation as the final common trigger for pressure overload hypertrophy. More recently, these mice have shown resistance to heart failure following chronic hypertrophic stimulus (Esposito *et al.*, 2002), suggesting that Gq-class specific inhibition is a novel strategy to prevent ventricular dysfunction in conditions of chronic hypertrophic stress.

## C. MANIPULATION OF CARDIAC GRK EXPRESSION

### 1. $\beta$ ARK1

The importance of  $\beta$ ARK1 (GRK2) in the cardiovascular system is clearly noted by the severe cardiac malformations and embryonic death observed



following  $\beta$ ARK1 gene ablation (Jaber *et al.*, 1996). The findings suggest a possible role for  $\beta$ ARK1 in the normal migration and differentiation of myocardial cells during heart development. Heterozygous  $\beta$ ARK1 knockout mice with 50% less  $\beta$ ARK1 expression and activity in myocardium have no developmental abnormalities (Rockman *et al.*, 1998b). Transgenic mice that overexpress  $\beta$ ARK1 in the heart due to the use of the  $\alpha$ MHC promoter have an attenuated response to catecholamine stimulation with desensitized  $\beta$ ARs (Koch *et al.*, 1995). This was a significant finding, as it represents the first demonstration that  $\beta$ ARK1 could cause the functional uncoupling of  $\beta$ ARs *in vivo*. Furthermore, these mice demonstrate that the upregulation of  $\beta$ ARK1 seen in human heart failure may have significance and contribute to the pathogenesis of ventricular dysfunction. Contractile responses to angiotensin II also are abrogated in  $\alpha$ MHC- $\beta$ ARK1 transgenic mice, suggesting that  $\beta$ ARK1 may be important in other receptor systems in the heart (Rockman *et al.*, 1996). Interestingly,  $\beta$ ARK1 overexpression has no effect on cardiac  $\alpha_{1B}$ AR signaling, demonstrating GRK-GPCR selectivity *in vivo* (Eckhart *et al.*, 2000).

## 2. Inhibition of $\beta$ ARK1

Since  $\beta$ ARK1 activity is increased in heart failure and appears to play a role in uncoupling of  $\beta$ ARs in the heart, we have studied the physiological consequences of  $\beta$ ARK1 inhibition. To do this, a specific peptide inhibitor consisting of the last 194 amino acids of the  $\beta$ ARK1 ( $\beta$ ARKct) was developed and studied (Koch *et al.*, 1993). The  $\beta$ ARKct contains the  $G_{\beta\gamma}$  binding domain and competes with endogenous  $\beta$ ARK1 for  $G_{\beta\gamma}$ -mediated membrane translocation, a process required for  $\beta$ ARK1 activation on activated GPCRs (Koch *et al.*, 1993). When the  $\beta$ ARKct was expressed in the hearts of transgenic mice under the control of the  $\alpha$ MHC promoter, cardiac physiology was altered in reciprocal fashion to that seen with  $\beta$ ARK1 overexpression (Koch *et al.*, 1995). The  $\beta$ ARKct mice have enhanced cardiac function at baseline and an augmented response to catecholamines (Koch *et al.*, 1995). Importantly, using a hybrid transgenic mouse strategy where  $\beta$ ARK1 overexpression and  $\beta$ ARKct expression occurred *in vivo* simultaneously, we have shown that the  $\beta$ ARKct is, indeed, inhibiting cardiac  $\beta$ ARK1 activity (Akhter *et al.*, 1999).

The phenotypes of the  $\beta$ ARK1 and the  $\beta$ ARKct mice are consistent with our hypothesis that this GRK plays a critical role in cardiac function and potentially in cardiac pathologies. Interestingly, heterozygous  $\beta$ ARK1 knockout mice also have a phenotype of enhanced cardiac function (Rockman *et al.*, 1998b), demonstrating that lowering  $\beta$ ARK1 expression or its activity can have profound *in vivo* effects on cardiac contractility. Moreover, hybrid mice that express the  $\beta$ ARKct in the heart and are heterogeneous for  $\beta$ ARK1 gene ablation have even-greater enhancement of cardiac function (Rockman *et al.*, 1998b).



In addition to heart failure, enhanced  $\beta$ ARK1 expression and activity has been shown to be indicative of several models of cardiac hypertrophy (Koch *et al.*, 2000). Enhanced  $\beta$ ARK1 activity in the hypertrophied heart has been shown to be responsible for the loss of  $\beta$ AR inotropic reserve seen in this pathological condition (Choi *et al.*, 1997). To study the inhibition of  $\beta$ ARK1 during hypertrophy, we used a novel transgenic mouse model. The  $\beta$ ARKct was targeted to the heart using the cardiac ankyrin repeat protein (CARP) promoter, which turns off during adulthood. However CARP belongs to a family of fetal genes, such as atrial natriuretic factor (ANF), that can be reactivated in adult ventricular myocardium by stress. CARP- $\beta$ ARKct mice lose  $\beta$ ARKct expression after 3 weeks of life and adult mice do not have enhanced contractility (Manning *et al.*, 2000). However, following induction of pressure overload hypertrophy, expression of the  $\beta$ ARKct is seen once again in the myocardium, resulting in improved  $\beta$ AR responsiveness and cardiac function (Manning *et al.*, 2000).

### 3. Cardiac Transgenic Studies with GRK3 and GRK5

Following the profound effects seen with  $\beta$ ARK1 manipulation in the hearts of transgenic mice, we studied the physiological consequences of GRK3 and GRK5 overexpression. These two GRKs are found normally in the heart but their overall role in cardiac signaling is not well understood, although GRK5 has been found to be upregulated in some animal models of heart failure (Ping *et al.*, 1997; Vinge *et al.*, 2001). Unlike  $\beta$ ARK1, both GRK3 and GRK5 homozygous knockout mice are viable with no overt cardiac phenotype (Wess, 2000). Overexpression of these GRKs in the heart has, however, led to unexpected and interesting results that have uncovered novel aspects of GRK regulation *in vivo* in the heart. GRK3 (also known as  $\beta$ ARK2) previously was thought to be an isozyme of  $\beta$ ARK1, since it is highly homologous and appeared to have the same *in vitro* GPCR activity (Benovic *et al.*, 1991; Freedman *et al.*, 1995). However, when  $\alpha$ MHC-GRK3 mice were generated and studied, there were no signaling alterations in the cardiac  $\beta$ AR system (Iaccarino *et al.*, 1998a). This was the first demonstration *in vivo* that GRK3 was different from  $\beta$ ARK1, with a unique GPCR specificity profile. Further investigation revealed that thrombin signaling in the heart was uncoupled in these mice, demonstrating that the thrombin receptors are *in vivo* substrates for GRK3 (Iaccarino *et al.*, 1998a). The difference in these GRKs may lie in the  $G_{\beta\gamma}$  binding regions, which is the area between GRK3 and  $\beta$ ARK1 that is the most divergent (Muller *et al.*, 1997). Thus, there may be selective GPCR-mediated translocation of these GRKs. In hybrid transgenic mice with different GRKs overexpressed along with the  $\alpha_{1B}$ AR, it was found that GRK3 is also the primary kinase for desensitization of this AR in the heart (Eckhart *et al.*, 2000).

GRK5, which is the second-highest expressing GRK in the heart, is not regulated by  $G_{\beta\gamma}$  and thus would be expected to have different receptor substrates in the heart. However, like  $\beta$ ARK1 overexpressing mice,  $\alpha$ MHC-GRK5 transgenic mice had severely blunted  $\beta$ AR inotropic responses *in vivo* in the heart, demonstrating that this GRK also could desensitize cardiac  $\beta$ ARs (Rockman *et al.*, 1996). These mice exhibited GPCR substrate selectivity, compared to  $\beta$ ARK1, as responses to angiotensin II were not altered, whereas this Gq-coupled receptor system was desensitized in mice overexpressing  $\beta$ ARK1 (Rockman *et al.*, 1996). GRK5 also has some activity against cardiac  $\alpha_{1B}$ ARs, again demonstrating a difference with  $\beta$ ARK1 (Eckhart *et al.*, 2000). The overall significance of the findings that GRK5 may be altered in heart failure is not clear at this time but obviously could have important implications.

### III. $\beta$ ARK1 Inhibition and Rescue of Murine Models of Heart Failure

One interesting area where this research has led us is to investigate whether inhibition of  $\beta$ ARK1 activity could be a novel therapeutic strategy for improving function of the failing heart. Over the last few years, this has become possible to study in the mouse, as murine models of cardiomyopathy have been described, many of which have important manifestations of the human condition. These models have been the result of a specific gene deletion in the mouse or cardiac-specific overexpression of a heart failure-inducing transgene. Powerful information can be generated by cross-breeding  $\alpha$ MHC- $\beta$ ARKct mice and various heart failure models to test the hypothesis that  $G_{\beta\gamma}$ - $\beta$ ARK1 inhibition could be beneficial. Simply studying the cardiac phenotype of these novel hybrid mice could give an answer and provide information on the role of  $\beta$ ARK1 and GRK activity in the pathogenesis of the various heart failure etiologies, specific for the different models. We have studied six different heart failure models with  $\beta$ ARKct mice and, for the most part, have seen overwhelming rescue. Table II summarizes our findings over the last 3–4 years using this novel genetic approach; some of the more-important findings are detailed in the next paragraph.

The first murine heart failure model rescued by any genetic manipulation was done by us with  $\beta$ ARKct animals (Rockman *et al.*, 1998a). The model of heart failure was due to the gene knockout of the muscle LIM protein (MLP), a cytoarchitectural protein and conserved regulator of myogenic differentiation (Rockman *et al.*, 1998a). The improvements made by the  $\beta$ ARKct in this model of dilated cardiomyopathy ( $MLP^{-/-}$ ) included restoring cardiac chamber dilation, increasing basal contractility, and enhancing  $\beta$ AR function (Table II). A second model of heart failure with different characteristics of disease and rescue was due to cardiac-targeted overexpression of calsequestrin (CSQ), a high-capacity calcium-binding protein (Harding *et al.*, 2001). These mice have much more-severe disease than the  $MLP^{-/-}$  animals and experience early mortality, as

TABLE II  
*Murine Models of Heart Failure and Rescue Status with Cardiac  $\beta$ ARKct Expression*

| Murine model  | Phenotypic change with $\beta$ ARKct   | Reference                     |
|---|--|-------------------------------|
| MLP <sup>-/-</sup> knockout   | Functional rescue with restored $\beta$ AR responsiveness                                    | Rockman <i>et al.</i> , 1998a |
| Transgenic cardiac CSQ overexpression   | Functional rescue and cardiac dimensions with improved survival                              | Harding <i>et al.</i> , 2001  |
| Transgenic cardiac expression of a mutant myosin heavy chain (HCM)                            | Functional rescue, prevention of hypertrophy, and dimensions and improved exercise tolerance | Freeman <i>et al.</i> , 2001  |
| Transgenic cardiac overexpression of MCP-1  | Prevention of hypertrophy  | Khouri <i>et al.</i> , 2002   |
| Transgenic cardiac overexpression of dominant-negative mutant of CREB (CREB <sub>A133</sub> ) | Restoration of $\beta$ AR signaling, no functional or mortality rescue                       | Eckhart <i>et al.</i> , 2002a |
| Transgenic cardiac overexpression of G $\alpha$ q   | No rescue  | Dorn <i>et al.</i> , 1999     |

[Abbreviations: MLP<sup>-/-</sup>, homozygous knockout of muscle LIM protein; CSQ, calsequestrin; MCP-1, monocyte chemotactic protein-1; CREB, cyclic AMP-responsive element binding protein.]

all mice are dead within 20 weeks of life. Thus, in this study, we were able to carry out a survival test.  $\beta$ ARKct expression nearly doubled the life-span of these heart-failure mice (Harding *et al.*, 2001). In addition to survival, the  $\beta$ ARKct prevented excessive ventricular deterioration and improved cardiac function (Harding *et al.*, 2001). Furthermore, combination of  $\beta$ -blocker therapy (metoprolol) and  $\beta$ ARKct expression was synergistic in improving survival (Harding *et al.*, 2001). These data are particularly interesting, given the clinical promise of  $\beta$ -blocker therapy in human heart failure (Packer *et al.*, 1996,2001). Studies from our laboratory have also shown that chronic  $\beta$ AR blockade by carvedilol decreases the expression of  $\beta$ ARK1 in the heart and reduces cardiac GRK activity (Iaccarino *et al.*, 1998b).

As detailed in Table II, cardiac  $\beta$ ARKct expression also has rescued other models of heart failure, including one with cardiac-targeted overexpression of a mutant form of the  $\alpha$ MHC gene (HCM) that is associated with human hypertrophic cardiomyopathy (Freeman *et al.*, 2001). Interestingly, expression of the  $\beta$ ARKct in the G $\alpha$ q mice had no effect on the Gq phenotype, unlike the  $\beta_2$ AR (Dorn *et al.*, 1999). In the Gq model of decompensated cardiac hypertrophy,

$\beta$ ARK1 is not upregulated, suggesting that the  $\beta$ ARKct is acting specifically to inhibit GRKs. However, the exact mechanism of the  $\beta$ ARKct may involve sequestration of  $G_{\beta\gamma}$  from other signaling pathways, such as those involved in the activation of phosphoinositide-3 kinase (PI3K) (Naga Prasad *et al.*, 2000,2001) and  $I_{K,Ach}$  channels (Clapham and Neer, 1997). The contribution of these other potential  $G_{\beta\gamma}$  effects to the salutary effects of  $\beta$ ARKct in heart failure remains to be determined. Finally, the therapeutic benefit of the  $\beta$ ARKct may involve enhanced signaling through other GPCRs such as angiotensin II receptors.

#### IV. Hypertension, the Adrenergic Pathway, and Heart Failure

The American Heart Association suggests that the presence of hypertension or high blood pressure in a patient doubles that person's risk for developing heart failure. In essential hypertension, elevated blood pressure has been associated with increased sympathetic output (Mark, 1990), suggesting that the catecholamines and associated adrenergic pathways may be involved in this pathology. It also implicates hypertension as a potential primary component in the development of heart failure. Indeed, studies have shown that agents that reduce blood pressure, no matter what the mechanism, all appear to eventually reverse hypertrophy (Testa *et al.*, 1996; Anker *et al.*, 1997).

We previously discussed the importance of Gq signaling in cardiac hypertrophy and heart failure. As was detailed, transgenic mice that express the GqI peptide inhibitor of Gq can prevent the development of hypertrophy and heart failure in a pressure-overload model (Akhter *et al.*, 1998; Esposito *et al.*, 2002). To study the potential contribution of the vascular system and its associated alterations in blood pressure to this hypertrophic response, we developed a line of transgenic mice that express GqI in vascular smooth muscle cells under the control of the SM22 $\alpha$  promoter (Keys *et al.*, 2002). Following chronic Gq agonist administration, we observed an attenuation of mean arterial blood pressure and an inhibition of cardiac hypertrophy in the transgenic mice with vascular GqI expression (Keys *et al.*, 2002). In contrast — and somewhat unexpectedly — when the GqI was expressed in the heart, neither hypertension nor hypertrophy was inhibited (Keys *et al.*, 2002). These findings suggest that, during hypertension, the vascular system is the principal determinant of cardiac hypertrophy, rather than direct stimulation of the heart itself.

Interestingly, impairment of the vascular  $\beta$ AR system has been shown in human and animal models of hypertension (Feldman, 1990; Brodde and Michel, 1992). More specifically, elevations in  $\beta$ ARK1 expression have been found in lymphocytes of hypertensive patients (Gros *et al.*, 1997,1999). Recently, we generated transgenic mice that express  $\beta$ ARK1 in the vascular smooth muscle, again using the SM22 $\alpha$  promoter (Eckhart *et al.*, 2002b). These mice display attenuated vascular  $\beta$ AR signaling, an increase in mean blood pressure, and

develop cardiac hypertrophy (Eckhart *et al.*, 2002b). This again implicates the adrenergic system in hypertrophy – and, in particular, cardiovascular  $\beta$ ARK1 activity — which also proposes a link between hypertension and heart failure.

#### IV. Conclusion

The development of transgenic mouse models has provided a broader understanding of the physiological impact of individual proteins during heart failure. Overall, through our efforts detailed herein and those from other laboratories around the world, there are currently at least 75 genetically altered mouse models available to study the role of particular signaling systems in the heart (Chu *et al.*, 2002). This review has focused on the adrenergic signaling pathway under normal conditions and during heart failure. It is evident that transgenic mice have given us insight into the role of adrenergic system in the heart that otherwise would not have been possible. In the future, we hope that this knowledge may yield novel therapeutic interventions for the treatment of cardiac disease. In fact, adenoviral-mediated delivery of the  $\beta$ ARKct and  $\beta_2$ AR to larger animal models of heart failure has resulted in beneficial effects (White *et al.*, 2000; Shah *et al.*, 2001; Tevæarai *et al.*, 2002), suggesting that gene-therapy strategies may, indeed, target these AR abnormalities in heart failure in the coming years and offer new hope to patients suffering from this disease of epidemic proportions.

#### REFERENCES

- Adams JW, Sakata Y, Davis MG, Sah VP, Wang Y, Liggett SB, Chien KR, Brown JH, Dorn GW II** 1998 Enhanced Gq signaling: a common pathway mediates cardiac hypertrophy and apoptotic heart failure. *Proc Natl Acad Sci USA* 95:10140–10145
- Akhter SA, Milano CA, Shotwell KF, Cho MC, Rockman HA, Lefkowitz RJ, Koch WJ** 1997 Transgenic mice with cardiac overexpression of  $\alpha_{1B}$ -adrenergic receptors. *J Biol Chem* 272:21253–21259
- Akhter SA, Luttrell LM, Rockman HA, Iaccarino G, Lefkowitz RJ, Koch WJ** 1998 Targeting the receptor-Gq interface to inhibit *in vivo* pressure overload myocardial hypertrophy. *Science* 280:574–577
- Akhter SA, Eckhart AD, Rockman HA, Shotwell KF, Lefkowitz RJ, Koch WJ** 1999 *In vivo* inhibition of elevated myocardial  $\beta$ -adrenergic receptor kinase activity in hybrid transgenic mice restores normal  $\beta$ -adrenergic signaling and function. *Circulation* 100:648–653
- Anker SD, Chau TP, Ponikowski P, Harrington D, Swan JW, Kox WJ, Poole-Wilson PA, Coats AJS** 1997 Hormonal changes and catabolic/anabolic imbalance in chronic heart failure and their importance for cardiac cachexia. *Circulation* 96:526–534
- Asai K, Yang GP, G YJ, Takagi G, Bishop S, Ishikawa Y, Shannon RP, Wagner TE, Vatner DE, Homey CJ, Vatner SF** 1999  $\beta$ -adrenergic receptor blockade arrests myocyte damage and preserves cardiac function in the transgenic G $\alpha_s$  mouse. *J Clin Invest* 104:551–558
- Baker AJ, Redfern CH, Harwood MD, Simpson PC, Conklin BR** 2001 Abnormal contraction caused by expression of Gi coupled receptor in transgenic model of dilated cardiomyopathy. *Am J Physiol* 280:H1653–H1659

- Benovic JL, Onorato JJ, Arriza JL, Stone WC, Lohse M, Jenkins NA, Gilbert DJ, Copeland NG, Caron MG, Lefkowitz RJ** 1991 Cloning, expression and chromosomal localization of  $\beta$ -adrenergic receptor kinase 2. *J Biol Chem* 266:14939–14946
- Bristow MR, Ginsburg R, Umans V, Fowler M, Minobe W, Rasmussen R, Zera P, Menlove R, Shah P, Jamieson S, Stinson EB** 1982 Decreased catecholamine sensitivity and  $\beta$ -adrenergic receptor density in failing human hearts. *N Engl J Med* 307:205–211
- Bristow MR, Herschberger RE, Port JD, Rasmussen R** 1989  $\beta_1$  and  $\beta_2$  adrenergic receptor mediated adenyl cyclase stimulation in non-failing and failing human ventricular myocardium. *Mol Pharmacol* 35:395–399
- Bristow MR, Minobe W, Reynolds MV, Port JD, Rasmussen R, Ray PE, Feldman AM** 1993 Reduced  $\beta_1$  receptor messenger RNA abundance in the failing human heart. *J Clin Invest* 92:2737–2745
- Brodde OE** 1993 Beta-adrenoceptors in cardiac disease. *Pharmacol Ther* 60:405–443
- Brodde O-E, Michel MC** 1992 Adrenergic receptors and their signal transduction mechanisms in hypertension. *J Hypertens* 10:S133–S145
- Caron MG, Lefkowitz RJ** 1993 Catecholamine receptors: structure, function and regulation. *Recent Prog Horm Res* 48:277–290
- Chein KR** 1999 Stress pathways and heart failure. *Cell* 98:555–558
- Choi D-J, Koch WJ, Hunter JJ, Rockman HA** 1997 Mechanism for  $\beta$ -adrenergic receptor desensitization in cardiac hypertrophy is increased  $\beta$ -adrenergic receptor kinase. *J Biol Chem* 272:17223–17229
- Chu G, Haghghi K, Kranias EG** 2002 From mouse to man: understanding heart failure through genetically altered mouse models. *J Card Failure* 8:S432–S449
- Clapham DE, Neer EJ** 1997 G protein beta gamma subunits. *Annu Rev Pharmacol Toxicol* 37:167–203
- Cohn JN, Levine TB, Olivari MT, Garberg V, Lura D, Francis GS, Simon AB, Rector T** 1984 Plasma norepinephrine as a guide to prognosis in patients with chronic congestive heart failure. *N Engl J Med* 311:819–823
- D'Angelo DD, Sakata Y, Lorenz JN, Boivin GP, Walsh RA, Liggett SB, Dorn GW II** 1997 Transgenic  $G\alpha_q$  overexpression induces cardiac contractile failure in mice. *Proc Natl Acad Sci USA* 94:8121–8126
- Dorn GW II, Tepe NM, Lorenz JN, Davis MG, Koch WJ, Liggett SB** 1999 Low and high  $\beta_2$  adrenergic receptors differentially affect cardiac hypertrophy and function in  $G\alpha_q$  overexpressing mice. *Proc Natl Acad Sci USA* 96:6400–6405
- Du X-J, Autelitano DJ, Dilley RJ, Wang B, Dart AM, Woodcock EM** 2000  $\beta_2$  adrenergic receptor overexpression exacerbates development of heart failure after aortic stenosis. *Circulation* 101:71–77
- Eckhart AD, Duncan SJ, Penn RB, Benovic JL, Lefkowitz RJ, Koch WJ** 2000 Hybrid transgenic mice reveal *in vivo* specificity of G protein-coupled receptor kinases in the heart. *Circ Res* 86:43–50
- Eckhart AD, Fentzke RC, Lepore J, Lang R, Lin H, Lefkowitz RJ, Leiden JM, Koch WJ** 2002a Inhibition of  $\beta$ ARK1 and restoration of myocardial  $\beta$ -adrenergic signaling in a mouse model of dilated cardiomyopathy induced by CREB<sub>A133</sub> expression. *J Mol Cell Cardiol* 34:669–677
- Eckhart AD, Ozaki T, Tevaearai H, Rockman HA, Koch WJ** 2002b Vascular targeted overexpression of a G protein coupled receptor kinase 2 in transgenic mice attenuates  $\beta$ -adrenergic receptor signaling and increases resting blood pressure. *Mol Pharmacol* 61:749–758
- Engelhardt S, Hein L, Weismann F, Lohse MJ** 1999 Progressive hypertrophy and heart failure in  $\beta_1$  adrenergic receptor transgenic mice. *Proc Natl Acad Sci USA* 96:7059–7064

- Epstein SE, Braunwald E** 1966 The effect of  $\beta$ -adrenergic blockade on patterns of urinary sodium excretion. Studies in normal subjects and in patients with heart disease. *Ann Intern Med* 65:20–27
- Esler M, Kaye D, Malber G, Esler D, Jennings G** 1997 Adrenergic nervous system in heart failure. *Am J Cardiol* 80:7L–14L
- Esposito G, Rapacciuolo A, Prasad SV, Takaoka H, Thomas SA, Koch WJ, Rockman HA** 2002 Genetic alterations that inhibit *in vivo* pressure – overload hypertrophy prevent cardiac dysfunction despite increased wall stress. *Circulation* 105:85–92
- Feldman AM, Cates AE, Veazey WB, Hershberger RE, Bristow MR, Baughman KL, Baumgartner WA, Van Dop E** 1988 Increase of the 40000 mol wt pertussis toxin substrate (G protein) in the failing human heart. *J Clin Invest* 82:189–197
- Feldman RD** 1990 Defective venous  $\beta$ -adrenergic response in borderline hypertensive subjects is corrected by a low sodium diet. *J Clin Invest* 85:647–652
- Freedman NJ, Liggett SB, Drachman DE, Pei G, Caron MG, Lefkowitz RJ** 1995 Phosphorylation and desensitization of the human  $\beta_1$  adrenergic receptor: involvement of G protein-coupled receptor kinase and cAMP dependent protein kinase. *J Biol Chem* 270:17953–17961
- Freeman K, Olsson MC, Iaccarino G, Bristow MR, Lefkowitz RJ, Kranias EL, Koch WJ, Leinwand LA** 2001 Alterations in cardiac adrenergic signaling and calcium cycling differentially affect the progression of cardiomyopathy. *J Clin Invest* 107:967–974
- Gaudin C, Ishikawa Y, Wight DC, Mahavi V, Nada-Ginnard B, Wagner TE, Vatner DE, Homcy CJ** 1995 Overexpression of G $\alpha$  protein in the hearts of transgenic mice. *J Clin Invest* 95:1676–1683
- Geng YJ, Ishikawa Y, Vatner DE, Wagner TE, Bishop SP, Vatner SF, Homcy CJ** 1999 Apoptosis of cardiac myocytes in G $\alpha$  transgenic mice. *Circ Res* 84:34–42
- Green SA, Cole G, Jacinto M, Innis M, Liggett SB** 1993 A polymorphism of the human  $\beta_2$  adrenergic receptor within the fourth transmembrane domain alters ligand binding and functional properties of the receptor. *J Biol Chem* 268:23116–23121
- Gros R, Benovic JL, Tan C, Feldman RD** 1997 G protein-coupled receptor kinase activity is increased in hypertension. *J Clin Invest* 99:2087–2093
- Gros R, Tan C, Chorzyczewski J, Kelvin DJ, Benovic JL, Feldman RD** 1999 G protein coupled receptor kinase expression in hypertension. *Clin Pharmacol Ther* 65:545–551
- Grossman W, Jones D, McLaurin LP** 1975 Wall stress and patterns of hypertrophy in the human left ventricle. *J Clin Invest* 56:56–64
- Harding VB, Jones LR, Lefkowitz RJ, Koch WJ, Rockman HA** 2001 Cardiac  $\beta$ ARK1 inhibition prolongs survival and augments  $\beta$ -blocker therapy in a mouse model of severe heart failure. *Proc Natl Acad Sci USA* 98:5809–5814
- Iaccarino G, Rockman HA, Shotwell KF, Tomhave ED, Koch WJ** 1998a Myocardial targeted overexpression of G protein-coupled receptor kinase 3 in transgenic mice: evidence for *in vivo* selectivity of GRKs. *Am J Physiol* 275:H1298–H1306
- Iaccarino G, Tomhave ED, Lefkowitz RJ, Koch WJ** 1998b Reciprocal *in vivo* regulation of myocardial G protein-coupled receptor kinase expression by  $\beta$ -adrenergic receptor stimulation and blockade. *Circulation* 98:1783–1789
- Iaccarino G, Keys JR, Rapacciuolo A, Shotwell KF, Lefkowitz RJ, Rockman HA, Koch WJ** 2001 Regulation of myocardial  $\beta$ ARK1 expression in catecholamine induced cardiac hypertrophy in transgenic mice overexpressing  $\alpha_{1B}$  adrenergic receptors. *J Am Cell Cardiol* 38:541–545
- Inglese J, Freedman NJ, Koch WJ, Lefkowitz RJ** 1993 Structure and mechanism of the G protein-coupled receptor kinases. *J Biol Chem* 268:23735–23738



- Jaber M, Koch WJ, Rockman HA, Smith B, Bond RA, Sulik KK, Ross J Jr, Lefkowitz RJ, Caron MG, Giros B** 1996 Essential role of  $\beta$ -adrenergic kinase 1 in cardiac development and function. *Proc Natl Acad Sci USA* 93:12974–12979
- Jain M, Lim CC, Nagata K, Davis KM, Milstone DS, Liao R, Mortensen RM** 2001 Targeted inactivation of  $G\alpha_i$  does not alter cardiac function of beta adrenergic sensitivity. *Am J Physiol* 280:H569–H575
- Keys JR, Greene EA, Koch WJ, Eckhart AD** 2002 Gq-coupled receptor agonists mediate cardiac hypertrophy via the vasculature. *Hypertension* 40:660–666
- Khouri SJ, Binkley P, Koch WJ, Kolattukudy P** 2002 Myocardial overexpression of the cardiac  $\beta$ -adrenergic receptor kinase-1 inhibitor ( $\beta$ ARKI) delays the development of cardiomyopathy induced by myocardial expression of monocyte chemo-tactic protein-1 (MCP-1). *J Am Coll Cardiol* 39:I-164 (abstract)
- Koch WJ, Inglese J, Stone WC, Lefkowitz RJ** 1993 The binding site for the  $\beta\gamma$  subunits of heterotrimeric G proteins on the  $\beta\beta$ -adrenergic receptor kinase. *J Biol Chem* 268:8256–8260
- Koch WJ, Rockman HA, Samama P, Hamilton RA, Bond RA, Milano C, Lefkowitz RJ** 1995 Cardiac function in mice overexpressing the  $\beta$ -adrenergic receptor kinase or a  $\beta$ ARK inhibitor. *Science* 268:1350–1353
- Koch WJ, Lefkowitz RJ, Rockman HA** 2000 Functional consequences of altering myocardial adrenergic receptor signaling. *Ann Rev Physiol* 63:237–260
- Liggett SB, Tepe NM, Lorenz JL, Canning AM, Jantz TD, Mitarai S, Yatani A, Dorn GW II** 2000 Early and delayed consequences of  $\beta_2$  adrenergic receptor overexpression in mouse hearts. *Circulation* 101:1707–1714
- Manning BS, Shotwell K, Mao L, Rockman HA, Koch WJ** 2000 Physiological induction of a  $\beta$ -adrenergic receptor kinase inhibitor transgene preserves  $\beta$ -adrenergic responsiveness in pressure overload cardiac hypertrophy. *Circulation* 102:2751–2757
- Mark AL** 1990 Regulation of sympathetic nerve activity in mild human hypertension. *J Hypertens* 8:S67–S75
- Maurice JD, Hata JA, Shah AS, White DC, McDonald PH, Dolber PC, Wilson KH, Lefkowitz RJ, Glower DD, Koch WJ** 1999 Enhancement of cardiac function after adenoviral mediated *in vivo* intracoronary  $\beta_2$  adrenergic receptor gene delivery. *J Clin Invest* 104:21–29
- Milano CA, Allen LF, Rockman HA, Dolber PC, McMinn TR, Chein KR, Johnson TD, Bond RA, Lefkowitz RJ** 1994a Enhanced myocardial function in transgenic mice overexpressing the  $\beta_2$  adrenergic receptor. *Science* 264:582–586
- Milano CA, Dolber PC, Rockman HA, Bond RA, Venable ME, Allen LF, Lefkowitz RJ** 1994b Myocardial expression of a constitutively active  $\alpha_{1B}$  adrenergic receptor in transgenic mice induces cardiac hypertrophy. *Proc Natl Acad Sci USA* 91:10109–10113
- Molkentin JD, Dorn GW II** 2001 Cytoplasmic signaling pathways that regulate cardiac hypertrophy. *Ann Rev Physiol* 63:391–426
- Muller S, Straub A, Lohse MJ** 1997 Selectivity of  $\beta$ -adrenergic receptor kinase 2 for G protein  $\beta\gamma$  subunits. *FEBS Lett* 401:25–29
- Naga Prasad SV, Esposito G, Mao L, Koch WJ, Rockman HA** 2000  $G\beta\gamma$  dependent phosphoinositide 3-kinase activation in hearts with *in vivo* pressure overload hypertrophy. *J Biol Chem* 275:4693–4698
- Naga Prasad SV, Barak LS, Rapacciuolo A, Caron MG, Rockman HA** 2001 Agonist-dependent recruitment of phosphoinositide 3-kinase to the membrane by  $\beta$ -adrenergic receptor kinase 1. A role in receptor sequestration. *J Biol Chem* 276:18953–18959
- Packer M, Bristow MR, Cohn JN, Colucci WS, Fowler MB, Gilbert EM, Shusterman NH** 1996 The effect of carvedilol on morbidity and mortality in patients with chronic heart failure. *N Engl J Med* 334:1349–1355

- Packer M, Coats AJ, Fowler MB, Katus HA, Krum H, Mohacsi P, Rouleau JL, Tendera M, Castaigne A, Roecker EB, Shultz MK, DeMets DL** 2001 Effect of carvedilol on survival in severe chronic heart failure. *N Engl J Med* 344:1651–1658
- Ping P, Anzai T, Gao M, Hammond HK** 1997 Adenylyl cyclase and G protein receptor kinase expression during development of heart failure. *Am J Physiol* 273:H707–H717
- Podlowski S, Wenzel K, Luther HP, Muller J, Bramlage P, Bauman G, Feliz SB, Speer A, Hetzer R, Kopke K, Hoehe MR, Wallukat G** 2000  $\beta_1$  adrenoreceptor gene variations: a role in idiopathic dilated cardiomyopathy? *J Mol Med* 78:87–93
- Redfern CH, Coward P, Degtyarev MY, Lee EK, Kwa AT, Hennighausen L, Bujard H, Fishman GI, Conklin BR** 1999 Conditional expression and signaling of a specifically designed Gi coupled receptor in transgenic mice. *Nature Biotechnol* 17:165–169
- Rockman HA, Choi D-J, Rahman NU, Akhter SA, Lefkowitz RJ, Koch WJ** 1996 Receptor specific *in vivo* desensitization by the G protein coupled receptor kinase 5 in transgenic mice. *Proc Natl Acad Sci USA* 93:9954–9959
- Rockman HA, Chein KR, Choi D-J, Iaccarino G, Hunter JJ, Ross J Jr, Lefkowitz RJ, Koch WJ** 1998a Expression of a  $\beta$ -adrenergic receptor kinase 1 inhibitor prevents the development of heart failure in gene-targeted mice. *Proc Natl Acad Sci USA* 95:7000–7005
- Rockman HA, Choi D-J, Akhter SA, Jobe M, Goris B, Lefkowitz RJ, Caron MF, Koch WJ** 1998b Control of myocardial contractile function by the level of  $\beta$ -adrenergic receptor kinase 1 in gene targeted mice. *J Biol Chem* 273:18180–18184
- Rockman HA, Koch WJ, Lefkowitz RJ** 2002 Seven-transmembrane-spanning receptors and heart function. *Nature* 415:206–212
- Rohrer DK** 1998 Physiological consequences of  $\beta$ -adrenergic receptor disruption. *J Mol Med* 76:764–772
- Rohrer DK, Desai KH, Jasper JR, Stevens ME, Regula DP, Barsh GS, Berstein D, Kobilka BK** 1996 Targeted disruption of the mouse  $\beta_1$  adrenergic receptor gene: developmental and cardiovascular effects. *Proc Natl Acad Sci USA* 93:7375–7380
- Rohrer DK, Schauble EH, Desai KH, Kobilka BK, Berstein D** 1998 Alterations in dynamic heart rate control in the  $\beta_1$  adrenergic receptor knockout mouse. *Am J Physiol* 274:H1184–H1193
- Shah AS, White DC, Emani S, Kypson AP, Lilly RE, Wilson K, Glower DD, Lefkowitz RJ, Koch WJ** 2001 *In vivo* ventricular gene delivery of a  $\beta$ -adrenergic receptor kinase inhibitor to the failing heart reverses cardiac dysfunction. *Circulation* 103:1311–1316
- Small KM, Wagoner LE, Levin AM, Kardia SL, Liggett SB** 2002 Synergistic polymorphisms of  $\beta_1$ - and  $\alpha_2C$ -adrenergic receptors and the risk of congestive heart failure. *N Engl J Med* 347:1135–1142
- Testa M, Yeh M, Lee P, Fanelli R, Loperfido F, Berman JW, LeJemtel TH** 1996 Circulating levels of cytokines and their endogenous modulators in patients with mild to severe congestive heart failure due to coronary artery disease or hypertension. *J Am Coll Cardiol* 28:964–971
- Tevaearai HT, Eckhart AD, Walton GB, Keys JR, Wilson K, Koch WJ** 2002 Myocardial gene transfer and overexpression of  $\beta_2$ -adrenergic receptors potentiates the functional recovery of unloaded failing hearts. *Circulation* 106:124–129
- Turki J, Lorenz JN, Green SA, Donnelly ET, Jacinto M, Liggett SB** 1996 Myocardial signaling defects and impaired cardiac function of a human  $\beta$ -adrenergic receptor polymorphism expressed in transgenic mice. *Proc Natl Acad Sci USA* 93:10483–10488
- Ungerer M, Bohm M, Elce JS, Erdmann E, Lohse MJ** 1993 Altered expression of  $\beta$ -adrenergic receptor kinase and  $\beta_1$ -adrenergic receptors in the failing human heart. *Circulation* 87:454–463

- Ungerer M, Parruti G, Bohm M, Puzicha M, DeBlasi A, Erdmann E, Lohse MJ** 1994 Expression of  $\beta$ -arrestins and  $\beta$ -adrenergic receptor kinases in the failing human heart. *Circ Res* 74:206–213
- Vinge LE, Oie E, Anderson Y, Groggaard HK, Andersen G, Attramadal H** 2001 Myocardial distribution and regulation of GRK and  $\beta$ -arrestin isoforms in congestive heart failure in rats. *Am J Physiol* 281:H2490–H2499
- Wagoner LE, Craft LL, Singh B, Suresh DP, Zengel PW, McGuire N, Abraham WT, Chenier TC Dorn GW II, Liggett SB** 2000 Polymorphisms of the  $\beta_2$ -adrenergic receptor determine exercise capacity in patients with heart failure. *Circ Res* 86:834–840
- Wess J** 2000 Physiological roles of G-protein-coupled receptor kinases revealed by gene-targeting technology. *Trends Pharmacol Sci* 21:364–366
- White DC, Hata JA, Shah AS, Glower DD, Lefkowitz RJ, Koch WJ** 2000 Preservation of myocardial  $\beta$ -adrenergic receptor delays the development of heart failure following myocardial infarction. *Proc Natl Acad Sci USA* 97:5428–5433
- Xiao RP, Avdonin P, Zhou YY, Cheng H, Akhter SA, Eschenhagen T, Lefkowitz RJ, Koch WJ, Lakatta EG** 1999 Coupling of  $\beta_2$ -adrenoceptor to Gi proteins and its physiological relevance in murine cardiac myocytes. *Circ Res* 84:43–52
- Zusic MJ, Chalothorn D, Hellard D, Deighan C, McGee A, Daly CJ, Waugh DJ, Ross SA, Gaivin RJ, Morehead AJ, Thomas JD, Plow EF, McGrath JC, Piascik MT, Perez DM** 2001 Hypotension, autonomic failure and cardiac hypertrophy in transgenic mice overexpressing the  $\alpha_{1B}$  adrenergic receptor. *J Biol Chem* 276:13738–13743